

## New Pyrrole Alkaloids from *Solanum sodomaeum*

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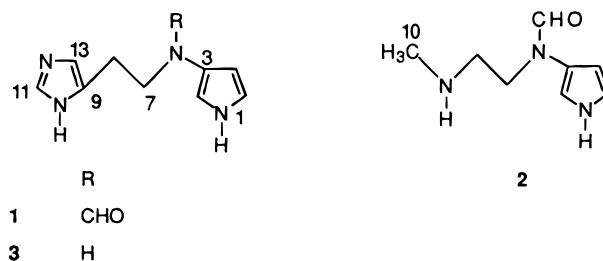
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Two new pyrrole alkaloids, solsodomine A and B, were isolated from the fresh berries of *Solanum sodomaeum* L., collected from the Libyan desert. The structures of these compounds were established by 2D-NMR, including <sup>15</sup>N NMR spectroscopy and chemical degradation. Solsodomine A (**1**) shows activity against *Mycobacterium intracellulare*. This is the first report of pyrrole alkaloids from the genus *Solanum*.

*Solanum* is one of the largest genera in the plant kingdom. It includes about 1400 species widely distributed throughout the world.<sup>1,2</sup> *Solanum* has been extensively studied for its steroidal alkaloids, nitrogen-containing compounds, and isoprenoids.<sup>3</sup> *Solanum* alkaloids exhibit an interesting range of bioactivity. Solapartine, solapalmitine, and solapalmitenine show tumor inhibitory activity against human nasopharynx carcinoma (KB).<sup>4</sup> Solamargine possesses potent cytotoxic activity to human hepatocyte (Hep3B) and normal skin fibroblasts.<sup>5</sup> Other *Solanum* alkaloids show significant antifungal and antihepatotoxic activities.<sup>6,7</sup> *Solanum sodomaeum* L. (Solanaceae) is an annual herb, widely distributed in the Libyan desert.<sup>8</sup> A topical formulation containing purified glycoalkaloids from *S. sodomaeum* L. has proven effective in the treatment of malignant human skin tumors; basal cell carcinomas, squamous cell carcinomas, and the benign tumors; keratoses, and keratoacanthomas.<sup>9</sup> Another study of the mixture of glycoalkaloids extracted from the title plant indicated its antineoplastic activity against Sarcoma 180 in mice.<sup>10</sup> This paper represents the first study of the less polar nonsteroidal alkaloids from *S. sodomaeum* and the first report for pyrrole alkaloids from the genus *Solanum*.

The alkaloid-containing extract of fresh berries of *S. sodomaeum* L. was fractionated into the less polar (CHCl<sub>3</sub>) and the more polar (EtOAc) fractions. Unlike the EtOAc fraction, the CHCl<sub>3</sub> fraction did not give positive Lieberman–Burchard, Salkowski, or Molish's tests, indicating the absence of steroidal glycosides.<sup>11</sup> TLC screening of the CHCl<sub>3</sub> fraction indicated the presence of alkaloids by a positive Dragendorff's test.<sup>11</sup> Fractionation of the CHCl<sub>3</sub> extract on silica gel 60 afforded two new pyrrole alkaloids, named solsodomine A (**1**) and B (**2**). The FTIR spectra of **1** showed absorptions at 3465 and 1657 cm<sup>-1</sup>, suggesting the presence of NH and *N*-formyl functionalities. The CIMS spectra of **1** displayed a molecular ion peak at *m/z* 204, consis-



tent with the molecular formula C<sub>10</sub>H<sub>12</sub>ON<sub>4</sub> and the presence of seven degrees of unsaturation. The proton singlet at  $\delta$  9.40 is correlated to a methine carbon resonating at  $\delta$  179.1, indicating the presence of an *N*-formyl group (Table 1). The downfield methylene triplet ( $\delta$  4.44) is correlated by the HMQC experiment to the nitrogenated methylene carbon at  $\delta$  48.6 (C-7). The C-7 methylene group displayed COSY coupling to another methylene triplet ( $\delta$  2.93), which is correlated in the HMQC spectrum to the methylene carbon at  $\delta$  28.6 (C-8). The presence of the pyrrole functionality was indicated from the two COSY-coupled doublet of doublets resonating at  $\delta$  6.05 and 6.87, correlating to the two methine carbons at  $\delta$  109.3 and 125.3 (C-4 and C-5, respectively). Both protons ( $\delta$  6.05 and 6.87) show long-range COSY couplings to a broad proton singlet resonating at  $\delta$  6.75, correlated to the methine carbon at  $\delta$  131.9 (C-2). The imidazole moiety was suggested from the two long-range COSY-coupled singlets resonating at  $\delta$  7.47 and 6.57, which are correlated to the methine carbons absorbing at  $\delta$  134.2 and 116.6 (C-11 and C-13 respectively). Analysis of the HMBC data of **1** further supported the proposed structure (Figure 1). The <sup>3</sup>*J*-HMBC correlations of H-14 with C-3 and C-7 supported its location on N-6. The <sup>3</sup>*J*-HMBC correlations of H-2 with C-4 and C-5 further supported the pyrrole ring entity. The same proved true for the imidazole ring with <sup>3</sup>*J*-HMBC correlations between H-11, C-9, and C-13. Additional confirmation was obtained from the analysis of <sup>1</sup>H–<sup>15</sup>N HMBC data (Figure 1, Table 1). The downfield nitrogen resonating at  $\delta$  225.2 (N-10) show <sup>3</sup>*J*-HMBC correlations to H-8 and H-13, in addition to a <sup>2</sup>*J*-HMBC correlation to H-11. The nitrogen resonating at  $\delta$  190.8 (N-12) shows <sup>2</sup>*J*-HMBC correlations to H-11 and H-13. The nitrogen (N-1) resonating at  $\delta$

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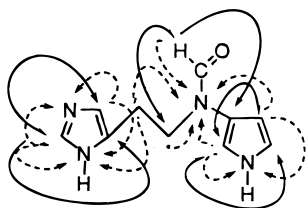
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**Table 1.**  $^{13}\text{C}$  and  $^1\text{H}$  NMR Spectral Data of Compounds **1** and **2**<sup>a</sup>

position	<b>1</b>			<b>2</b>	
	$\delta_{\text{C}}$	$\delta_{\text{H}}$	$\delta_{\text{N}}$	$\delta_{\text{C}}$	$\delta_{\text{H}}$
1		5.86, br s	165.8		5.87, br s
2	131.9, d	6.75, br s		132.9, d	7.03, br s
3	130.4, s			132.8, s	
4	109.3, d	6.05 dd (4.0, 2.5)		109.8, d	6.22 dd (4.1, 2.5)
5	125.3, d	6.87 dd (4.0, 2.4)		125.7, d	6.96 dd (4.1, 2.4)
6			165.6		
7	48.6, t	4.44 t (7.2)		51.1, t	4.42 t (7.1)
8	28.6, t	2.93 t (7.3)		50.3, t	3.84 t (7.2)
9	133.5, s				u
10		5.86, br s	225.2	31.8, q	3.41, s
11	134.2, d	7.47, s			
12			190.8		
13	116.6, d	6.57, s			
N <sub>6</sub> -formyl	179.1, d	9.40, s		179.8, d	9.42, s

<sup>a</sup> In  $\text{CDCl}_3$ , at 400 MHz for  $^1\text{H}$ , 100 MHz for  $^{13}\text{C}$ , and 50 MHz for  $^{15}\text{N}$ . Carbon multiplicities were determined by the DEPT 135° experiment; s = quaternary, d = methine, t = methylene, q = methyl carbons, u = unobserved; coupling constants ( $J$ ) are in Hz.

**Figure 1.** Important  $^1\text{H}$ - $^{13}\text{C}$  (solid lines) and  $^1\text{H}$ - $^{15}\text{N}$  (dashed lines) HMBC correlations of **1**.

165.8 shows  $^2J$ -HMBC correlations to H-2 and H-5, as well as a  $^3J$ -HMBC correlation to H-4. Similarly, the nitrogen (N-6) absorbing at  $\delta$  165.6 showed  $^2J$ -HMBC correlations to H-7 and the downfield *N*-formyl proton singlet, in addition to  $^3J$ -HMBC correlations to H-2, H-4, and H-8. The downfield shifting of the aliphatic N-6 is attributed to the deshielding effect of the *N*-formyl moiety. Acid hydrolysis of **1** afforded **3** which has similar  $^1\text{H}$  NMR spectral data except for the absence of the *N*-formyl signal.

The CIMS spectra of solsodamine B (**2**) displayed a molecular ion peak at  $m/z$  168  $[\text{M} + \text{H}]^+$ , consistent with the molecular formula  $\text{C}_8\text{H}_{13}\text{ON}_3$  and the presence of four degrees of unsaturation. The FTIR and  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra (Table 1) of **2** showed distinct similarity to those of **1** except for the lack of the imidazole moiety and the emergence of a new *N*-methyl group. The proton singlet resonating at  $\delta$  3.41 correlated to the methyl carbon at  $\delta$  31.8 (C-10). The downfield shift of the methylene proton triplet resonating at  $\delta$  3.84 and correlated to the methylene carbon at  $\delta$  50.3 is attributed to the nitrogenated C-8.

Solsodamine A (**1**) shows moderate activity against *Mycobacterium intracellulare* with an MIC of 10  $\mu\text{g}/\text{mL}$ . Compounds **1** and **2** did not show *in vitro* antimalarial activity against *Plasmodium falciparum* W2 and D6 clones, cytotoxic activity to Vero cells, or antifungal activity against *Cryptococcus neoformans* and *Candida albicans*.

This is the first report for pyrrole alkaloids in the genus *Solanum*; however, they have been previously

reported in the family Solanaceae from *Nicotiana bigelovii* and *Nicotiana setchellii*.<sup>2</sup>

## Experimental Section

**General Experimental Procedures.** Melting points were determined on a Thomas-Hoover capillary melting point apparatus and are uncorrected. UV spectra were acquired on a Perkin-Elmer Lambda 3B UV/vis spectrophotometer. The IR spectra were recorded on an ATI Mattson Genesis Series FTIR spectrophotometer. The  $^1\text{H}$  and  $^{13}\text{C}$  NMR spectra were recorded in  $\text{CDCl}_3$  on a Bruker DRX-400 spectrometer. The  $^{15}\text{N}$  NMR was recorded on a Bruker DRX-500 spectrometer. The LRCIMS spectra were obtained at the University of Kansas on a Fisons/VG Autospec Q mass spectrometer. TLC analysis was carried out on precoated silica gel G<sub>254</sub> (Merck) or aluminum oxide ALOX-100 UV<sub>254</sub> (Macherey-Nagel) plates. For column chromatography, Baker silica gel 60, 40  $\mu\text{m}$  was used.

**Plant Material.** Fresh *S. sodomaenum* L. berries were collected near Tripoli during spring 1994. The plant was identified by Dr. A. El-Gadi, Department of Botany, Al Faateh University, Tripoli, Libya. A voucher specimen (94Trip-3) was deposited at Pharmacognosy Department, Mansoura University, Egypt.

**Extraction and Isolations.** Fresh *S. sodomaenum* L. berries (4.5 kg) were minced, and the formed paste was extracted twice each with 2 L of 95% hot ethanol and concentrated under reduced pressure. The semi-solid ethanolic extract (750 g) was treated with 1 L of 5% aqueous tartaric acid and filtered. The filtrate was extracted twice, each with 1 L of  $\text{CHCl}_3$ . The acid-soluble fraction was alkalized to pH 9, using concentrated  $\text{NH}_4\text{OH}$ , and then extracted with  $\text{CHCl}_3$  (1 L  $\times$  3) followed by EtOAc (1 L  $\times$  3). The chloroform fraction was subjected to repeated column chromatography on silica gel 60 using *n*-hexane and gradient elution with EtOAc. The alkaloid-containing fractions were further subjected to preparative TLC on  $\text{Al}_2\text{O}_3$ , using cyclohexanes-EtOAc (2:1) to afford compounds **1** (85.5 mg,  $R_f$  0.45) and **2** (2.2 mg,  $R_f$  0.71).

**Solsodamine A (1):** yellow needles from EtOH; mp 118–119 °C; UV  $\lambda_{\text{max}}$  ( $\log \epsilon$ ) (MeOH) 280 (3.08), 320 (3.06), 348 (2.93) nm; IR  $\nu_{\text{max}}$  ( $\text{CHCl}_3$ ) 3465, (NH), 3006–2931, 1657 ( $\text{N}-\text{CH}=\text{O}$ )  $\text{cm}^{-1}$ ;  $^1\text{H}$  and  $^{13}\text{C}$  NMR data, see Table 1; LR-CIMS  $m/z$  (fragment) 204.0 ( $\text{M}^+$ , 15) (calcd for  $\text{C}_{10}\text{H}_{12}\text{ON}_4$ , 204.10), 190.0 (95), 176 (25), 162 (65), 94 (55), 81 (30), 69 (20).

**Acid Hydrolysis of 1.** Forty mg of **1** was dissolved in 1 mL of EtOH, and 5 mL of 6 N HCl was added. The reaction mixture was heated for 120 min at 80 °C. The pH was adjusted to 7, using dilute  $\text{NH}_4\text{OH}$ , and then the reaction mixture was extracted with  $\text{CHCl}_3$  (15 mL  $\times$  2). The  $\text{CHCl}_3$  solution was evaporated and subjected to preparative TLC on silica gel G<sub>254</sub> using  $\text{CHCl}_3$ -MeOH (9:1) to afford compound **3** (3.8 mg,  $R_f$  0.32).

**Solsodamine B (2):** colorless needles from EtOH; mp 112–113 °C; UV  $\lambda_{\text{max}}$  ( $\log \epsilon$ ) (MeOH) 271 (3.10), 317 (3.01), 342 (2.03) nm; IR  $\nu_{\text{max}}$  ( $\text{CHCl}_3$ ) 3455, (NH), 2995–2930, 1655 ( $\text{N}-\text{CH}=\text{O}$ )  $\text{cm}^{-1}$ ;  $^1\text{H}$  and  $^{13}\text{C}$  NMR data, see Table 1; LR-CIMS  $m/z$  (fragment) 168.0  $[(\text{M} + \text{H})^+]$  (calcd for  $\text{C}_8\text{H}_{14}\text{ON}_3$ , 168.11), 124.0 (100), 79 (15), 43 (50).

**Compound 3:** yellow needles from EtOH; mp 122–123 °C; UV  $\lambda_{\text{max}}$  ( $\log \epsilon$ ) (MeOH) 266 (2.99), 319 (3.11),

341 (2.71) nm; IR  $v_{\max}$  (CHCl<sub>3</sub>) 3455, (NH), 3010–2950 cm<sup>-1</sup>; <sup>1</sup>H NMR data (400 MHz, CDCl<sub>3</sub>)  $\delta$  6.42 (1H, brs, H-2), 6.10 (1H, dd (4.0, 2.1), H-4), 6.53 (1H, dd (4.0, 2.2), H-5), 3.97 (2H, t (7.1), H7), 2.96 (2H, t (7.1), H-8), 7.43 (1H, s, H-11), 6.56 (1H, s, H-13).

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